

Guidelines for the Blood Transfusion and Tissue Transplantation Services in the UK

Annex 4. Redundant component specifications

This section contains information for reference regarding the following redundant components:

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A4.1. Granulocytes, Apheresis

Important

This is a redundant component and the specification is provided for reference only.

A component prepared from anticoagulated blood, which is separated into components by a suitable apheresis machine with retention of granulocytes as the major cellular product, suspended in a portion of the plasma. The remaining elements may be returned to the donor.

A4.1.1. Technical information

- The component is not leucocyte depleted.
- The component contains red cells and requires compatibility testing.
- Granulocytes may be collected by a variety of apheresis systems using different protocols. Since yields may vary, each procedural protocol must be fully validated, documented and specifications set accordingly.
- Cytomegalovirus (CMV) seronegative granulocytes should be considered for CMV seronegative recipients.
- The component must not be agitated during storage.
- The component must be irradiated before use.
- Granulocytes, Apheresis should be transfused through a 170–200 µm filter.

A4.1.2. Labelling

For general guidelines, see chapter 6.6.

The following shall be included on the label:

(* = in eye-readable and UKBTS approved barcode format)

- Granulocytes, Apheresis* and volume
- the blood component producer's name*
- the donation number*
- the ABO group*
- the RhD group stated as positive or negative*
- the date of collection
- the expiry date and time*
- the temperature of storage
- the statement 'Do not agitate'
- the blood pack lot number*

- the name, composition and volume of the anticoagulant solution

In addition, the following statements should be made:

“INSTRUCTION

Always check patient/component compatibility/identity

Inspect pack and contents for signs of deterioration or damage

Risk of adverse reaction/infection”

A4.1.3. Storage

For general guidelines, see chapter 6.7.

- Granulocytes, apheresis should be used as soon as possible after their preparation. If storage is unavoidable, provided the component has been prepared using a closed system, the component should be stored, without agitation, at a core temperature of $22 \pm 2^\circ\text{C}$ and used within 24 hours of collection.

A4.1.4. Testing

In addition to the mandatory and other tests required for blood donations described in chapter 9, all tested for the parameters shown in Table A4.1 shall meet the specified values.

Table A4.1: Granulocytes, Apheresis – additional tests

Parameter	Specification	Frequency of test
Volume	Within locally defined nominal volume range (≤ 500 mL)	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Total granulocyte count	$>1 \times 10^{10}$ /unit	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)

A4.1.5. Transportation

For general guidelines, see chapter 6.11.

- Containers for transporting Granulocytes, Apheresis should be equilibrated at room temperature before use. During transportation the temperature of the component must be kept as close as possible to the recommended storage temperature and, on receipt, unless intended for immediate therapeutic use, the component should be transferred to storage at a core temperature of $22 \pm 2^\circ\text{C}$.
- Plastic overwraps should be removed prior to storage.

A4.2. Convalescent Plasma (COVID-19), FFP, Leucocyte Depleted

Important

This is a redundant component and the specification is provided for reference only.

Plasma that has been obtained from whole blood or by apheresis from donors who have recovered from COVID-19 infection, for treatment of patients with COVID-19. The plasma contains less than 2.5×10^6 leucocytes per component and has been rapidly frozen to a temperature that will maintain the activity of labile coagulation factors.

A4.2.1. Technical information

- Donations of whole blood where the bleed time exceeded 15 minutes are not suitable for the production of plasma components for direct clinical use.
- Plasma can be selected from male or female donors. Female donors must be screened and negative for HLA/HNA antibodies, as a TRALI risk reduction measure. Plasma should only be selected as CP for treatment of patients with COVID-19 if it is validated to contain a minimum concentration of SARS-CoV-2 antibody levels according to national clinical guidelines.
- When manufactured from whole blood the plasma should be separated before the red cell component is cooled to its storage temperature. Greater FVIII yields will be obtained when the plasma is separated as soon as possible after venepuncture and rapidly frozen to -25°C or below.
- The method of preparation should be validated to ensure there is no evidence of significant activation at 24 hours shelf life, with minimum cellular contamination. The production process should be validated to ensure that components meet the specified limits for FVIII concentration. If plasma collected for CP were to be re-manufactured for any other purpose these procedures must be fully validated and in accordance with the specification of the alternative component.
- Component samples collected for the quality monitoring assessment of FVIII should be from an equal mix of group O and non-O donations due to the difference in FVIII levels between ABO blood groups.
- Convalescent Plasma (COVID-19), FFP, Leucocyte Depleted (LD) should be administered through a CE marked transfusion set.

A4.2.2. Labelling

For general guidelines, see chapter 6.6.

The following shall be included on the label:

(* = in eye-readable and UKBTS approved barcode format)

- Convalescent Plasma (COVID-19), FFP, Leucocyte Depleted* and volume
- the blood component producer's name*
- the donation number and, if divided, sub-batch number*
- the ABO group*
- the RhD group stated as positive or negative*
- the date of collection
- the expiry date of the frozen component*
- the temperature of storage
- the blood pack lot number*
- a warning that the component must be used within four hours of thawing if maintained at $22 \pm 2^{\circ}\text{C}$, or up to a maximum of 24 hours of thawing if stored at $4 \pm 2^{\circ}\text{C}$.
- the name, composition and volume of the anticoagulant.

In addition, the following statements should be made:

"INSTRUCTION

Always check patient/component compatibility/identity

Inspect pack and contents for signs of deterioration or damage

Risk of adverse reaction/infection"

A4.2.3. Storage

For general guidelines, see chapter 6.7.

- The component should be stored at a core temperature of -25°C or below for a maximum of 36 months.
- Although a storage temperature below -25°C improves the preservation of labile coagulation factors, lower temperatures increase the fragility of plastic. Particular care must be taken when handling such packs.
- The component should be thawed in a waterbath or other equipment designed for the purpose, within a vacuum-sealed overwrap bag according to a validated procedure. The optimal temperature at which the component should be thawed is 37°C ; temperatures between 33°C and 37°C are acceptable.
- Protocols must be in place to ensure that the equipment is cleaned daily and maintained to minimise the risk of bacterial contamination. After thawing, and at the time of administration, the content should be inspected to ensure that no insoluble cryoprecipitate is visible and that the container is intact.
- Once thawed, the component must not be refrozen and should be transfused as soon as possible. If delay is unavoidable, the component may be stored and should be used within 4 hours if maintained at $22 \pm 2^{\circ}\text{C}$ or up to a maximum of 24 hours if stored at $4 \pm 2^{\circ}\text{C}$.
- Transfusion of Convalescent Plasma (COVID-19), FFP, LD should be completed within 4 hours of issue out of a controlled temperature environment.

A4.2.4. Testing

In addition to the mandatory and other tests required for blood donations described in chapter 9, and leucocyte counting (see chapter 6.3 and chapter 7.1.1), a minimum of 75% of those components tested for the parameters shown in Table A4.2 shall meet the specified values with the exception of FVIII:C.

Table A4.2: Convalescent Plasma (COVID-19), FFP, LD – additional tests

Parameter	Specification	Frequency of test
Volume	Stated volume $\pm 10\%$	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Total protein	≥ 50 g/L	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Platelet count (3,6)	$< 30 \times 10^9$ /L	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Red cell count (3)	$< 6 \times 10^9$ /L	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
FVIII (4,5)	Mean ≥ 0.70 IU/mL	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Leucocyte count (1,2)	$< 2.5 \times 10^6$ /unit	As per chapter 6.3 and chapter 7.1.1 (but see [2] below)

Notes on Table A4.2

1. Methods validated for counting low numbers of leucocytes must be used.
2. 90% units should have less than 2.5×10^6 leucocytes and more than 99% of units should contain less than 5×10^6 leucocytes, both with 95% confidence.
3. Pre-freeze in starting component.
4. Units tested and found to have < 0.3 IU/mL should not be issued for transfusion.
5. A minimum of 90% of those components tested should have ≥ 0.50 IU/mL.
6. Units tested and found to have a platelet count $> 100 \times 10^9$ /L should not be issued for transfusion.

A4.2.5. Transportation

For general guidelines, see chapter 6.11.

Every effort should be made to maintain the core storage temperature during transportation. Unless the component is to be thawed and used straightaway, it should be transferred immediately to storage at the recommended temperature.

A4.3. Convalescent Plasma (COVID-19), FFP, for Neonates and Infants, Leucocyte Depleted

Important

This is a redundant component and the specification is provided for reference only.

Convalescent Plasma (COVID-19), FFP, for Neonates and Infants, Leucocyte Depleted (LD) is plasma that has been obtained from whole blood or by apheresis from donors who have recovered from COVID-19 infection, for treatment of patients with COVID-19. The plasma contains less than 2.5×10^6 leucocytes per component.

Using a closed system, the component may be subdivided into approximately equal volumes and rapidly frozen to a temperature that will maintain the activity of labile coagulation factors.

A4.3.1. Technical information

- Chapter 7.7 provides general guidance on the requirements for components for use in neonates and infants under 1 year.
- Donations of whole blood where the bleed time exceeded 15 minutes are not suitable for the production of plasma components for direct clinical use.
- The component should be free from clinically significant irregular blood group antibodies including high-titre anti-A and anti-B. Testing for CMV antibodies is not required.
- Plasma can be selected from male or female donors. Female donors must be screened and negative for HLA/HNA antibodies, as a TRALI risk reduction measure. Plasma should only be selected as CP for treatment of patients with COVID-19 if it is validated to contain a minimum concentration of SARS-CoV-2 antibody levels according to national clinical guidelines.
- When manufactured from whole blood the plasma should be separated before the red cell component is cooled to its storage temperature. Greater FVIII:C yields will be obtained when the plasma is separated as soon as possible after venepuncture and rapidly frozen to -25°C or below.
- The method of preparation should be validated to ensure there is no evidence of significant activation at 24 hours shelf life, with minimum cellular contamination. The production process should be validated to ensure that components meet the specified limits for FVIII:C concentration. If plasma collected for CP were to be re-manufactured for any other purpose these procedures must be fully validated and in accordance with the specification of the alternative component.
- Component samples collected for the quality monitoring assessment of FVIII:C should be from an equal mix of group O and non-O donations due to the difference in FVIII levels between ABO blood groups.
- Convalescent Plasma (COVID-19), FFP, for Neonates and Infants, LD should be transfused through a CE marked transfusion set.

A4.3.2. Labelling

For general guidelines, see chapter 6.6.

The following shall be included on the label:

(* = in eye-readable and UKBTS approved barcode format)

- Convalescent Plasma (COVID-19), FFP, for Neonates and Infants, Leucocyte Depleted* and volume
- the blood component producer's name*
- the donation number and, if divided, sub-batch number *
- the ABO group*
- the RhD group stated as positive or negative*
- the date of collection
- the expiry date of the frozen component*
- the temperature of storage
- the blood pack lot number*
- a warning that the component should be used within 4 hours of thawing if maintained at $22 \pm 2^{\circ}\text{C}$ or up to a maximum of 24 hours of thawing if stored at $4 \pm 2^{\circ}\text{C}$
- the name, composition and volume of the anticoagulant.

In addition, the following statements should be made:

“INSTRUCTION

Always check patient/component compatibility/identity

Inspect pack and contents for signs of deterioration or damage

Risk of adverse reaction/infection”

A4.3.3. Storage

For general guidelines, see chapter 6.7.

- The component should be stored at a core temperature of -25°C or below for a maximum of 36 months.
- Although a storage temperature below -25°C improves the preservation of labile coagulation factors, lower temperatures increase the fragility of plastic. Particular care must be taken when handling such packs.
- The component should be thawed in a waterbath or other equipment designed for the purpose, within a vacuum-sealed overwrap bag according to a validated procedure. The optimal temperature at which the component should be thawed is 37°C ; temperatures between 33°C and 37°C are acceptable.
- Protocols must be in place to ensure that the equipment is cleaned daily and maintained to minimise the risk of bacterial contamination. After thawing, and at the time of administration, the content should be inspected to ensure that no insoluble cryoprecipitate is visible and that the container is intact.

- Once thawed, the component must not be refrozen and should be transfused as soon as possible. If delay is unavoidable, the component may be stored and should be used within 4 hours if maintained at $22 \pm 2^{\circ}\text{C}$, or up to a maximum of 24 hours if stored at $4 \pm 2^{\circ}\text{C}$.
- Transfusion of Convalescent Plasma (COVID-19), FFP, for Neonates and Infants, LD should be completed within 4 hours of issue out of a controlled temperature environment.

A4.3.4. Testing

In addition to the mandatory and other tests required for blood donations described in chapter 9, and leucocyte counting (see chapter 6.3 and chapter 7.1.1), the component shall be free from clinically significant irregular blood group antibodies and high-titre anti-A and/or anti-B. Furthermore, a minimum of 75% of those components tested for the other parameters shown in Table A4.3 shall meet the specified values with the exception of FVIII:C.

Table A4.3: Convalescent Plasma (COVID-19), FFP, for Neonates and Infants, LD – additional tests

Parameter	Specification	Frequency of test
Volume	Stated volume $\pm 10\%$	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Total protein	≥ 50 g/L	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Platelet count (3,6)	$< 30 \times 10^9/\text{L}$	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Red cell count (3)	$< 6 \times 10^9/\text{L}$	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
FVIII (4,5)	Mean ≥ 0.70 IU/mL	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Leucocyte count (1,2)	$< 2.5 \times 10^6/\text{unit}$	As per chapter 6.3 and chapter 7.1.1 (but see [2] below)

Notes on Table A4.3

1. Methods validated for counting low numbers of leucocytes must be used.
2. 90% units should have less than 2.5×10^6 leucocytes and more than 99% of units should contain less than 5×10^6 leucocytes, both with 95% confidence.
3. Pre-freeze in starting component.
4. Units tested and found to have < 0.3 IU/mL should not be issued for transfusion.
5. A minimum of 90% of those components tested should have ≥ 0.50 IU/mL.
6. Units tested and found to have a platelet count $> 100 \times 10^9/\text{L}$ should not be issued for transfusion.

A4.3.5. Transportation

For general guidelines, see chapter 6.11.

Every effort should be made to maintain the core storage temperature during transportation. Unless the component is to be thawed and used straightaway, it should be transferred immediately to storage at the recommended temperature.

Last updated on 13 March 2024

A4.4. Cryoprecipitate, Methylene Blue Treated and Removed, Leucocyte Depleted

Important

This is a redundant component and the specification is provided for reference only.

This component is made for neonatal use:

- 7.7.12. Cryoprecipitate for Neonates and Infants, Pathogen Reduced, Leucocyte Depleted

A4.5. Plasma, Cryoprecipitate Depleted, Leucocyte Depleted

Important

This is a redundant component and the specification is provided for reference only.

The supernatant plasma removed during the preparation of Cryoprecipitate, Leucocyte Depleted (LD). The plasma from which the Plasma, Cryoprecipitate Depleted, LD was made contains less than 1×10^6 leucocytes per component and is derived from a previously tested donor.

A4.5.1. Technical information

- Donations of whole blood where the bleed time exceeded 15 minutes are not suitable for the production of plasma components for direct clinical use.
- Plasma should be selected from male donors or consideration should be given to screening female donors for HLA/HNA antibodies, as a TRALI risk reduction measure.
- Plasma, Cryoprecipitate Depleted, LD should be frozen to a core temperature of -25°C or below within 2 hours of separation from its Cryoprecipitate, Leucocyte Depleted.
- Plasma, Cryoprecipitate Depleted, LD should be transfused through a 170–200 μm filter.

A4.5.2. Labelling

For general guidelines, see chapter 6.6.

The following shall be included on the label:

(* = in eye-readable and UKBTS approved barcode format)

- Plasma, Cryoprecipitate Depleted, Leucocyte Depleted* and volume
- the blood component producer's name*
- the donation number*
- the ABO group*
- the RhD group stated as positive or negative*
- the date of collection
- the expiry date of the frozen component*
- the temperature of storage
- the blood pack lot number*
- a warning that the component must be used within 4 hours of thawing if maintained at $22 \pm 2^{\circ}\text{C}$, or 24 hours of thawing if stored at $4 \pm 2^{\circ}\text{C}$
- the name, composition and volume of the anticoagulant.

In addition, the following statements should be made:

“INSTRUCTION

Always check patient/component compatibility/identity

Inspect pack and contents for signs of deterioration or damage

Risk of adverse reaction/infection”

A4.5.3. Storage

For general guidelines, see chapter 6.7.

- The component should be stored at a core temperature of -25°C or below for a maximum of 36 months.
- Although a storage temperature below -25°C improves the preservation of labile coagulation factors, lower temperatures increase the fragility of plastic. Particular care must be taken when handling such packs.
- The component should be thawed in a waterbath or other equipment designed for the purpose, within a vacuum-sealed overwrap bag according to a validated procedure. The optimal temperature at which the component should be thawed is 37°C ; temperatures between 33°C and 37°C are acceptable.
- Protocols must be in place to ensure that the equipment is cleaned daily and maintained to minimise the risk of bacterial contamination. After thawing, the content should be inspected to ensure that no insoluble cryoprecipitate is visible and that the container is intact.
- Once thawed, the component must not be refrozen and should be transfused as soon as possible. If delay is unavoidable, the component may be stored and should be used within 4 hours if maintained at $22 \pm 2^{\circ}\text{C}$ or 24 hours if stored at $4 \pm 2^{\circ}\text{C}$, but it should be borne in mind that extended post-thaw storage will result in a decline in the content of labile coagulation factors.

A4.5.4. Testing

In addition to the mandatory and other tests required for blood donations described in chapter 9, and leucocyte counting (see chapter 6.3 and chapter 7.1.1), a minimum of 75% of those components tested for the other parameters shown in Table A4.5 shall meet the specified values.

Table A4.5: Plasma, Cryoprecipitate Depleted, LD – additional tests

Parameter	Specification	Frequency of test
Volume	Stated volume $\pm 10\%$	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Platelet count (2)	$< 30 \times 10^9/L$	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Red cell count (2)	$< 6 \times 10^9/L$	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Leucocyte count (1,2)	$< 2.5 \times 10^6/\text{unit}$	As per chapter 6.3 and chapter 7.1.1

Notes on Table A4.5

1. Methods validated for counting low numbers of leucocytes must be used.
2. Pre-freeze in starting component (fresh frozen plasma).

A4.5.5. Transportation

For general guidelines, see chapter 6.11.

Every effort should be made to maintain the core storage temperature during transportation. Unless the component is to be thawed and used straightaway, it should be transferred immediately to storage at the recommended temperature.

A4.6. Whole Blood, Leucocyte Depleted

Important

This is a redundant component and the specification is provided for reference only.

A unit of blood collected into an anticoagulant, containing less than 1×10^6 leucocytes.

A4.6.1. Technical information

- A unit of whole blood is collected in the UK from a suitable donor (see chapter 3 and chapter 5). The International Blood Pack specification contains 66.5 mL of anticoagulant and is suitable for the collection of 475 mL $\pm 10\%$.
- Whole Blood, Leucocyte Depleted (LD) should be administered through a CE/UKCA/UKNI marked transfusion set.

A4.6.2. Labelling

For general guidelines, see chapter 6.6.

The following shall be included on the label:

(* = in eye-readable and UKBTS approved barcode format)

- Whole Blood, Leucocyte Depleted* and volume
- the blood component producer's name*
- the donation number*
- the ABO group*
- the RhD group stated as positive or negative*
- the name, composition and volume of the anticoagulant solution
- the date of collection
- the expiry date*
- the temperature of storage
- the blood pack lot number.*

In addition, the following statements should be made:

“INSTRUCTION

Always check patient/component compatibility/identity

Inspect pack and contents for signs of deterioration or damage

Risk of adverse reaction/infection”

A4.6.3. Storage

For general guidelines, see chapter 6.7.

- The component may be stored for a maximum of 35 days at a core temperature of $4 \pm 2^\circ\text{C}$ if an adenine-supplemented anticoagulant is used, otherwise the maximum period of storage is 28 days at a core temperature of $4 \pm 2^\circ\text{C}$.
- Variation from the core temperature of $4 \pm 2^\circ\text{C}$ must be kept to a minimum during storage and restricted to any short period necessary for examining, labelling or issuing the component.
- Exceptionally, i.e. due to equipment failure at a Blood Centre, red cell components which have been exposed to a core temperature not exceeding 10°C and not less than 1°C may be released for transfusion provided that:
 - the component has been exposed to such a temperature change on one occasion only
 - the duration of the temperature excursion has not exceeded 5 hours
 - a documented system is available in each Blood Centre to cover such eventualities
 - adequate records of the incident are compiled and retained

A4.6.4. Testing

In addition to the mandatory and other tests required for blood donations described in chapter 9, and leucocyte counting (see chapter 6.3 and chapter 7.1.1), a minimum of 75% of those components tested for the parameters shown in Table A4.6 shall meet the specified values.

Table A4.6: Whole Blood, LD – additional tests

Parameter	Specification	Frequency of test
Volume (1)	475 mL $\pm 10\%$	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Haemolysis	$< 0.8\%$ of red cell mass	As per chapter 7.1.3
Haemoglobin content	≥ 40 g/unit	1% or as determined by statistical process control (if ≤ 10 components produced per month then test every available component)
Leucocyte count (2)	$< 1 \times 10^6$ /unit	As per chapter 6.3 and chapter 7.1.1

Notes on Table A4.6

1. After volume losses resulting from leucodepletion.
2. Methods validated for counting low numbers of leucocytes must be used.

A4.6.5. Transportation

For general guidelines, see chapter 6.11.

For red cell components, transit containers, packing materials and procedures should have been validated to ensure the component surface temperature can be maintained between 2°C and 10°C during transportation. Additionally:

- the validation exercise should be repeated periodically
- if melting ice is used, it should not come into direct contact with the components
- dead air space in packaging containers should be minimised
- as far as is practicable, transit containers should be equilibrated to their storage temperature prior to filling with components
- transport time normally should not exceed 12 hours

In some instances, it is necessary to issue red cell components that have not been cooled to their storage temperature prior to placing in the transit container. The transport temperature specified above is not applicable for such consignments.